

iRGD-modified lipid–polymer hybrid nanoparticles loaded with isoliquiritigenin to enhance anti-breast cancer effect and tumor-targeting ability

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Abstract: Isoliquiritigenin (ISL), a natural anti-breast cancer dietary compound, has poor delivery characteristics and low bioavailability. In order to promote the therapeutic outcome of ISL, a tumor-targeting lipid–polymer hybrid nanoparticle (NP) system modified by tumor-homing iRGD peptides has been developed. The hybrid NPs were prepared by a modified single-step nanoprecipitation method to encapsulate ISL. iRGD peptides were anchored on the surface by a postinsertion method (ISL-iRGD NPs). The stable lipid–polymer structure of ISL-iRGD NPs, with high encapsulation and loading efficiency, was confirmed. Compared to free ISL and non-iRGD-modified counterparts, ISL-iRGD NPs showed higher cytotoxicity and cell apoptosis against the different type of breast cancer cells. This was attributable to higher cellular accumulation mediated by the iRGD-integrin recognition and the nanoscale effect. More importantly, based on the active tumor-tissue accumulation by iRGD peptides and the prolonged in vivo circulation by the stealth nanostructure, ISL-iRGD NPs displayed higher tumor-growth inhibition efficiency in 4T1-bearing breast-tumor mouse models. Therefore, the constructed iRGD modified lipid–polymer hybrid NPs would provide a promising drug-delivery strategy to improve ISL in anti-breast cancer efficacy.

Keywords: isoliquiritigenin, iRGD, lipid–polymer hybrid NPs, breast cancer

Introduction

The morbidity of breast cancer has ranked first in female cancers for a long time worldwide. In some developed regions, breast cancer is still the second-commonest major cause of cancer death.^{1,2} Currently chemotherapy remains the irreplaceable method in treating breast cancer in clinic, yet it has limited efficacy and can produce some serious side effects for patients.^{3–6} On the other hand, seeking natural compounds in treating breast cancer has become a promising research area and attracted a lot of scholars, due to some compounds' high efficacy and minor side effects, such as curcumin, (–)-epigallocatechin gallate, and emodin.⁷ In very recent years, we have found a flavonoid, isoliquiritigenin (ISL; structure in Figure 1A), that is not only able to inhibit angiogenesis of breast cancer cells via the VEGF–VEGFR2 pathway⁸ and prevent mammary carcinogenesis through WIF1 demethylation,⁹ but can also chemosensitize breast cancer stem cells,¹⁰ serving as an autophagy inducer regulating chemoresistance-associated autophagy.¹¹ In addition, studies have shown that ISL can treat breast cancer via NFκB signaling and the PI3K–Akt pathway, inhibiting p38 expression and downregulating AA-metabolizing enzymes.^{12–14} This evidence has demonstrated that ISL can be a promising anti-breast cancer agent worthy of being

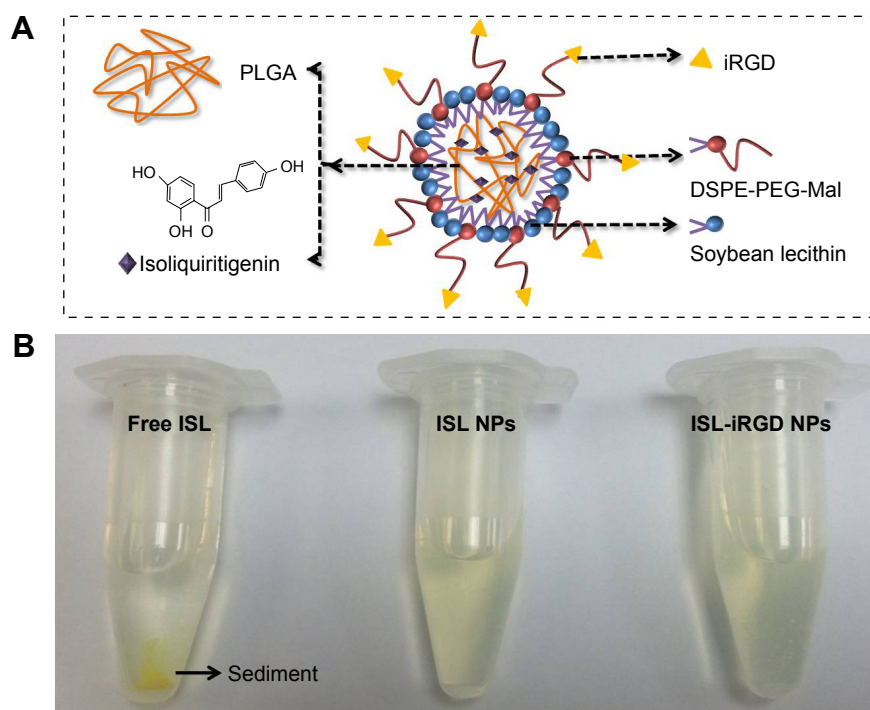


Figure 1 Image and structure of ISL-iRGD NPs and ISL.

Notes: Schematic representation of ISL-iRGD NPs (**A**); photographic image of different drugs (free ISL, ISL NPs and ISL-iRGD NPs) in aqueous solution (**B**).

Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles; PLGA, poly(lactic-co-glycolic acid); DSPE, 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine; PEG, polyethylene glycol; Mal, maleimide.

developed. However, currently its use in clinical application is limited by several factors, such as poor solubility, low bioavailability, high effective dose (50 mg/kg mouse body weight), and poor targeting ability (very low concentrations in mammary tissue).^{9,15} Therefore, dealing with these problems is a key process in its development.

Nanoparticles (NPs) are a typical nanodelivery system that can improve solubility, enhance bioavailability, and increase targeting distribution of a drug,¹⁶ while lipid-polymer NPs represent a promising delivery platform that combines the biomimetic advantages of liposomes and mechanical advantages of biodegradable polymeric NPs.^{17,18} In addition, modification of targeting peptides on the surface of the NPs serves as a smart design for enhancing targeting distribution.¹⁹ iRGD peptides, nine-unit cyclic (CRGDK/RGPD/EC) tumor-homing peptides with an RGD sequence were selected, since they bind preferentially to integrin $\alpha_v\beta_3$ receptors, $\alpha_v\beta_5$ integrin receptors and Nrp1 receptors. Firstly, the iRGD peptide can directly bind to $\alpha\beta$ integrin, which is overexpressed on the surface of cancer cells and tumor-vessel cells but not in normal vessel cells, and then proteolytically turns into CRGDK/R, exposing an active CendR motif at the C-terminus. Finally, the CendR motif binds with the permeability-regulating receptor Nrp1 in cancer cells.^{20–23}

Therefore, modified iRGD peptides on the surface of lipid-polymer NPs represent an effective design.

To the best of our knowledge, recent attempts to improve ISL delivery via nanosystems against cancer have been very limited and the outcomes not very satisfactory, ie, in the form of nanomaterial toxicity and poor targeting ability of existent NPs.^{24,25} Herein, iRGD-modified lipid-polymer NPs were used to encapsulate ISL (structure of NPs in Figure 1A). The basic characteristics, pharmacological efficacy, and targeting ability of the novel NPs were studied and are evaluated in this paper.

Materials and methods

Materials and reagents

The animal experiments were approved by the ethics committee of the University of Hong Kong (HKU, permit 3684-15), and we followed the animal-welfare guidelines and basic regulations of Laboratory Animal Unit in HKU. ISL (purity >97%) was bought from Thermo Fisher Scientific (Waltham, MA, US). 1,2-Distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-(maleimide[polyethylene glycol]_{2,000}) (DSPE-PEG_{2,000}-Mal) was purchased from Laysan Bio (Arab, AL, US). Poly(lactic-co-glycolic acid) (PLGA)-COOH (intrinsic viscosity [dL/g] 0.08, molecular

weight 5,000 Da) was obtained from Dalian Biotechnology (Nanjing, China). Lecithin was bought from Solarbio Science and Technology (Beijing, China). Coumarin 6 (C_6 ; laser grade) was obtained from Aladdin Reagent (Shanghai, China). 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) and Hoechst 33342 were purchased from Thermo Fisher Scientific. A fluorescein isothiocyanate (FITC)–annexin V apoptosis-detection kit used to investigate apoptosis activity was obtained from BD Biosciences (San Jose, CA, US). All reagent water was pretreated with the Milli-Q Plus system (EMD Millipore, Billerica, MA, US). All solvents, including dimethyl sulfoxide (DMSO), ethanol, acetonitrile, and methanol, were of chromatographic grade and used without further treatment.

Preparation of ISL-iRGD NPs

ISL-iRGD lipid–polymer NPs composed of a polymeric PLGA core coated with a layer of lipids and a PEG layer were prepared by modified single-step nanoprecipitation, based on a previous study.²⁶ In brief, DSPE-PEG–Mal (6 mg) and lecithin (4 mg) solution were dissolved in 0.5 mL alcohol and then added to 9 mL water. The DSPE-PEG–lecithin solution was stirred and heated to 65°C, to ensure all lipid was in liquid phase. Additionally, 1.2 mg of ISL and 10 mg of PLGA was dissolved in 0.5 mL of alcohol and acetonitrile, respectively. The PLGA solution mixed with ISL was then added to a prepared liquid-lipid solution dropwise under stirring at 700 rpm for 6 hours at room temperature. The remaining organic solvent was evaporated in a vacuum, and the nanosuspension was filtered with a 0.45 μ m filter to remove free molecules. Finally, iRGD peptides were modified on the surface of NPs by a postinsertion approach based on the mild conjugation between the Mal group on the PEG layer and the terminal sulfhydryl group of iRGD. iRGD (1 mg) was added to the ISL-loaded NP suspension for 30 minutes' shaking at room temperature.

Physical property characterization

The objective of studying property characterization was to evaluate the general physical properties and basic structures of the NPs, as well as to discover evidence to confirm whether the drugs were loaded into the NPs or not. NPs were characterized by particle size, ζ -potential, X-ray diffraction, infrared radiation, differential scanning calorimetry (DSC), and transmission electron microscopy (TEM). Particle size and ζ -potential were both detected by dynamic light scattering (Malvern Instruments, Malvern, UK) at 25°C. The morphology of NPs was detected by TEM (Tecnai G20; Thermo

Fisher Scientific) at operation voltage of 200 kV. Fourier-transform infrared spectra (Nicolet iS 50; Thermo Fisher Scientific) were employed to characterize the encapsulation of ISL in NPs. Thermodynamic properties of ISL-iRGD NPs were detected by diamond DSC (PerkinElmer, Waltham, MA, US). The crystallographic structures of NPs were characterized by X-ray diffraction (Bruker Optik GmbH, Ettlingen, Germany).

EE, LE, and stability analysis

Loading efficiency (LE) and encapsulation efficiency (EE) of ISL were determined with high-performance liquid-chromatography (HPLC) (Dionex Ultimate 3000; Thermo Fisher Scientific) equipped with a C_{18} column (250×4.6 mm). The mobile phase was methanol/water (70/30, v/v) with a detection wavelength of 306 nm. The flow rate was 1 mL/min. Methanol was used to disrupt the structure of NPs and dissolved ISL before it was analyzed by HPLC. Calculations of EE and LE were as follows:

$$EE (\%) = \frac{\text{Amount of ISL loaded}}{\text{Amount of ISL added}} \times 100\% \quad (1)$$

$$LE (\%) = \frac{\text{Amount of ISL loaded}}{\text{Amount of ISL added} + \text{polymer}} \times 100\% \quad (2)$$

In addition, change in particle size, ζ -potential, EE, and LE for 15 days during storage at 25°C was carried out as the stability evaluation.

Cell culture

Human breast cancer cell lines (MDA-MB231 and MCF7) and a mouse breast cancer cell line (4T1) were obtained from the American Type Culture Collection (Manassas, VA, US). Cells were cultured in medium (Dulbecco's Modified Eagle's Medium for MDA-MB231 and 4T1, Roswell Park Memorial Institute 1640 for MCF7) supplemented with 10% fetal bovine serum (Thermo Fisher Scientific) and 1% streptomycin and penicillin (Thermo Fisher Scientific). For all the in vitro experiments, free ISL was dissolved in DMSO in advance, and then diluted over 1,000-fold. All of the NPs or drugs were diluted by uncompleted medium (supplemented with 0.5% fetal bovine serum and 1% streptomycin and penicillin).

Cell-proliferation assays

MTT assay was used to evaluate the cell proliferation activity of different breast cancer cells. In brief, MDA-MB231, 4T1, and MCF7 cells were seeded onto the 96-well plates at a

density of 4×10^3 cells per well. Four groups (ISL-iRGD NPs, ISL NPs, free ISL, and blank NPs) were added to the wells with six duplicates for each concentration. After incubation for 24 hours, 20 μ L MTT solution (5 mg/mL) was added into each well for 4 hours' incubation. The MTT-added solution was then removed following the addition of 100 μ L of DMSO to each well and mixed thoroughly with pipette. Afterward, the samples were incubated at 37°C for 10 minutes and mixed. Absorbance was read at 540 nm.

Apoptosis assays

The FITC-annexin V apoptosis-detection kit was used to determine the effects of cancer-cell apoptosis. MDA-MB231 breast cancer cells were treated with the four drug groups (ISL-iRGD NPs, ISL NPs, free ISL, and blank NPs) with ISL concentration of 50 μ M for 16 hours. Both nonadherent and adherent cells were collected. Pretreatment procedures were carried out as per the instructions of the FITC-annexin V apoptosis-detection kit. Cells were analyzed with an LSR Fortessa analyzer (BD Biosciences) within 1 hour. Annexin V-FITC⁺/PI⁻ cells were considered early apoptotic cells, and annexin V-FITC⁺/PI⁺ cells were considered late apoptotic cells.

Cell-uptake analysis

Cellular uptake is an important index to estimate drug capacity to enter cells and drug concentration in cells.²⁷ C₆, a hydrophobic fluorescent dye, was used to label the NPs, and used to investigate the cell-uptake ability of different NPs qualitatively and quantitatively.²⁸ The preparation method for encapsulating C₆ was similar to ISL NPs.

Flow cytometry (FCM) was used for quantitative studies of the cellular uptake of NPs.^{29,30} MDA-MB231 breast cancer cells (8×10^5) were seeded onto the six-well plates. After 12 hours' incubation, the medium was removed and the different drug solutions added at certain concentrations of C₆. For the quantitative study, cells were incubated with C₆-iRGD NPs, C₆ NPs, and free C₆ with an equivalent concentration of C₆ (25 ng/mL) for 4 hours at 37°C. For dose-dependent study, different concentrations of C₆ (25, 12.5, 6.25, 3.125 ng/mL) were used for incubation with cells for 4 hours at 37°C. Furthermore, cells were incubated with C₆-iRGD NPs (25 ng/mL) at different time points (0.5, 1, 2, and 4 hours) for time-dependent study. For the energy-dependent study, cells were incubated with C₆-iRGD NPs (25 ng/mL) at different time points (0.5, 1, 2, and 4 hours) at 37°C and 4°C, respectively. Untreated cells were used as controls.

Cellular uptake of various ISL samples was observed by fluorescence microscopy. MDA-MB231 breast cancer cells (5×10^4) were seeded onto the six-well plates. After 24 hours' incubation, the completed medium was removed and different drug solutions at a concentration of C₆ (85 ng/mL) added. The control group was untreated cells. After 4 hours' incubation, the drug solutions were removed and washed twice with cold phosphate-buffered saline (PBS), fixed with 4% paraformaldehyde in PBS for 10 minutes, washed twice more with PBS, and then stained with Hoechst 33342. Intracellular Hoechst 33342 and C₆ were excited and observed by fluorescence microscopy.

In vivo antitumor efficacy

Primarily, 4-week-old female nude mice were used to set up the breast-tumor mouse models. 4T1 cells (4×10^5 per mouse) were injected into the mammary gland via subcutaneous inoculation. After the tumor volume had reached about 100 mm³, mice were assigned to four groups randomly: ISL-iRGD NPs, ISL NPs, free ISL (25 mg/kg mouse body weight), and blank NPs. Formulations were administered to mice once every 2 days by intraperitoneal injection for 20 days after disinfection. Mice in the control group were administered normal saline only. Tumor volumes and body weight of mice were recorded once every 2 days. The tumor volumes were calculated using the formula $V = W^2 \times L/2$. At the end of the experiment, mice were used anesthetic to death and collected tumors, livers, lungs, and kidneys. Hematoxylin and eosin (H&E) staining was used in studying the histological characteristics of the different tissues.

Biodistribution analysis

To visualize tumor accumulation of NPs in breast-tumor mouse models, Cy5, a near-infrared fluorescence probe, was used to label NPs based on the preparation method of ISL NPs. After injecting 4T1 breast cancer cells in the nude mice for 10 days, the mice were divided into Cy5-iRGD NPs, Cy5 NPs, and free Cy5 groups. Biodistribution analysis was conducted, described as follows. Firstly, mice were given Cy5 (35 μ g/kg) via intravenous (IV) injection. Secondly, mice were imaged using an IVIS Spectrum (PerkinElmer) in vivo imaging system after injecting the Cy5 at four time points (2, 6, 12, and 24 hours). Finally, tumors, lungs, hearts, spleens, and kidneys were collected and imaged.

Statistics

All data were presented as means \pm standard deviation (SD). All experimental results were confirmed in at least

three independent experiments under the same conditions. Statistical comparison was carried out with Student's *t*-test. $P < 0.05$ was considered statistically significant.

Results and discussion

Preparation and characterization of ISL-iRGD NPs

Based on the aforementioned procedures, ISL-loaded hybrid NPs composed of a polymeric PLGA core coated by a layer of lipids and PEG were prepared, and iRGD peptides were modified on the surface of NPs by postinsertion, with the structure shown in Figure 1A. With its significant improvement in water-solubility, the ISL-loaded NP suspension appeared as a homogeneous light-yellow solution, instead of lots of sediment in free ISL solution (Figure 1B).

The average size distribution of ISL-iRGD NPs was 137.2 ± 2.6 nm (relative SD 1.87%) and the ζ -potential -34.21 ± 1.23 mV (relative SD 3.59%) (Figure 2A and B). As is well known, NPs with small particle size less than 200 nm can accumulate in tumor cells via the enhanced permeability and retention (EPR) effect. Also, NPs with high absolute negative ζ -potential suggests satisfactory stability, as well as avoidance of particle aggregation and protein adsorption in blood circulation. Therefore, these NPs would be expected to show good stability and drug accumulation in tumor-tissue. Morphologically, the TEM images of ISL-iRGD NPs and blank NPs showed that distribution was uniform and the shape round and smooth (Figure 2C and D), which also indicated the loading procedure of ISL had not influenced NP formation.

The infrared spectra of ISL-iRGD NPs and other formulations were detected to confirm the structure of NPs. A strong peak at $1,228.494\text{ cm}^{-1}$ was detected only in the free ISL, mixed free ISL, and blank NPs, which indicated this peak was specific for ISL (Figure 2E). However, this peak disappeared from the ISL NPs, which means ISL was not detected on the surface of NPs and was loaded in the core. Moreover, the peaks of $2,921.771\text{ cm}^{-1}$, $2,854.272\text{ cm}^{-1}$, and $1,758.849\text{ cm}^{-1}$ were detected in mixed free ISL, blank NPs, and ISL NPs, respectively, but not free ISL itself, which is supplementary evidence to confirm ISL loading in the core of the NPs. In DSC or X-ray spectrum evaluation (Figure 2F and G), the spectrum profile or specific peaks of blank NPs were similar to ISL NPs; the same conclusion as for infrared-spectrum analysis. All the three spectra confirmed the same fact that ISL was encapsulated in the core of the NPs but not on the surface.

Drug-loading and -stability evaluation in vitro

The EE and LE of ISL-iRGD NPs in Milli-Q water were $90.8\% \pm 1.5\%$ and $4.13\% \pm 0.4\%$, respectively. The stability of ISL-iRGD NPs was tested for 15 days at 4°C . Results are shown in Table 1. The particle size of ISL-iRGD NPs changed from 138.97 ± 2.44 (day 1) to 134.48 ± 2.65 nm (day 15), but without significant difference ($P > 0.5$). In addition, the other three indices – ζ -potential, EE, and LE – all showed slight changes from day 1 to day 15. These findings indicated ISL-iRGD NPs were stable during the 15 days.

Cytotoxicity and apoptosis analysis of ISL-iRGD NPs in vitro

To evaluate the anti-breast cancer effects of ISL-iRGD NPs in vitro, MTT and double-staining apoptosis assays were conducted. Three typical breast cancer cell lines – MCF7, MDA-MB231, and 4T1 – were used in the MTT assay. These cell lines were incubated with four different formulations (blank NPs, free ISL, ISL NPs, and ISL-iRGD NPs) for 24 hours. For blank NPs, there was no obvious cytotoxicity against any cell lines, which was also evidence to eliminate the influence caused of the nanomaterials. For MCF7, ISL-iRGD NPs had better inhibition effects than free ISL at high concentrations ($> 12.5\text{ }\mu\text{M}$, $P < 0.05$) but they were not better than ISL NPs (Figure 3A). For MDA-MB231 and 4T1, ISL-iRGD NPs had better inhibition effects than ISL NPs and free ISL at all concentrations ($P < 0.05$), which means iRGD enhanced the efficacy of ISL NPs in cytotoxicity (Figure 3B and C). Moreover, this evidence indicates that MDA-MB231 and 4T1 were more sensitive to the ISL-iRGD NPs than MCF7.

Based on the high sensitivity of MDA-MB231 to the ISL-iRGD NPs in cytotoxicity assays, MDA-MB231 was selected to evaluate the apoptosis-inducing efficacy of different formulations, and the results are shown in Figure 3D and Figure S1. After incubation for 16 hours, the apoptosis rate of free ISL and ISL NPs was similar ($P > 0.05$), which indicated the nanoscale of ISL may not have contributed to the efficacy of inducing MDA-MB231-cell apoptosis. However, ISL-iRGD NPs presented higher rates of apoptosis (about 40%) than the other four groups, which suggested the stronger efficacy may have been caused by the tumor-homing iRGD peptides ($P < 0.05$).

Cell-uptake ability of ISL-iRGD NPs

As previously mentioned, iRGD is a tumor-homing peptide that shows positive targeting ability for cancer cells. Moreover, the average size of the ISL-iRGD NPs was about

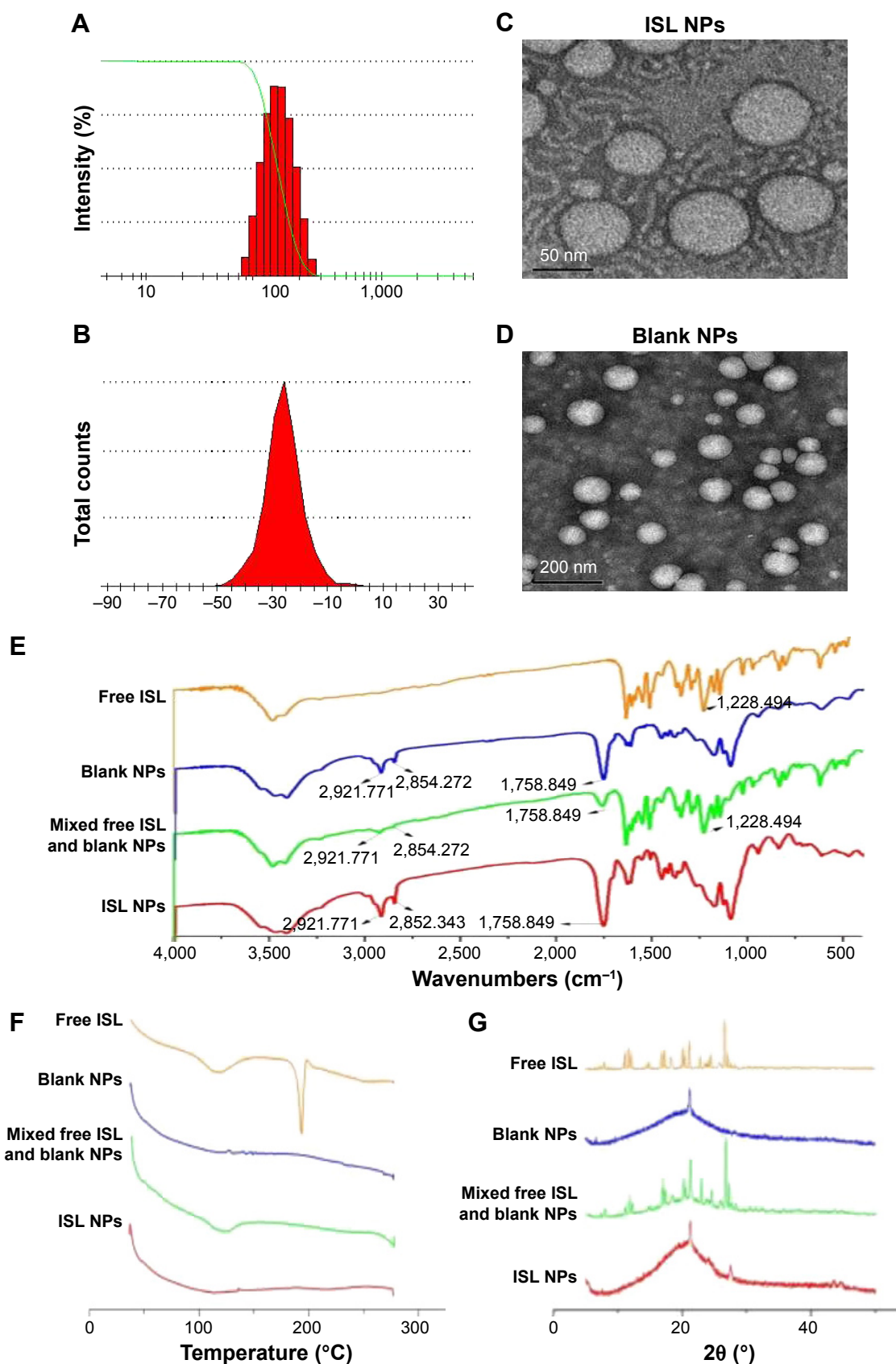


Figure 2 Characteristics of ISL-iRGD NPs.

Notes: Particle-size distribution (A), ζ -potential distribution (B), transmission electron microscopy (C [ISL-iRGD NPs], magnification $\times 200,000$; D [blank NPs]; magnification $\times 50,000$), infrared radiation (E), differential scanning calorimetry (F), and X-ray diffraction (G) of different polymer drug-delivery systems and free drug.

Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles.

Table 1 Stability of ISL-iRGD lipid-polymer NPs over 15 days

	Particle size	ζ -Potential	EE	LE
Day 1	138.97 \pm 2.44	-36.15 \pm 0.74	90.4% \pm 1.8%	4.06% \pm 0.38%
Day 15	134.48 \pm 2.65	-31.91 \pm 1.98	87.1% \pm 1.2%	3.91% \pm 0.4%

Note: All data were presented as mean \pm standard deviation.

Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles; EE, encapsulation efficiency; LE, loading efficiency.

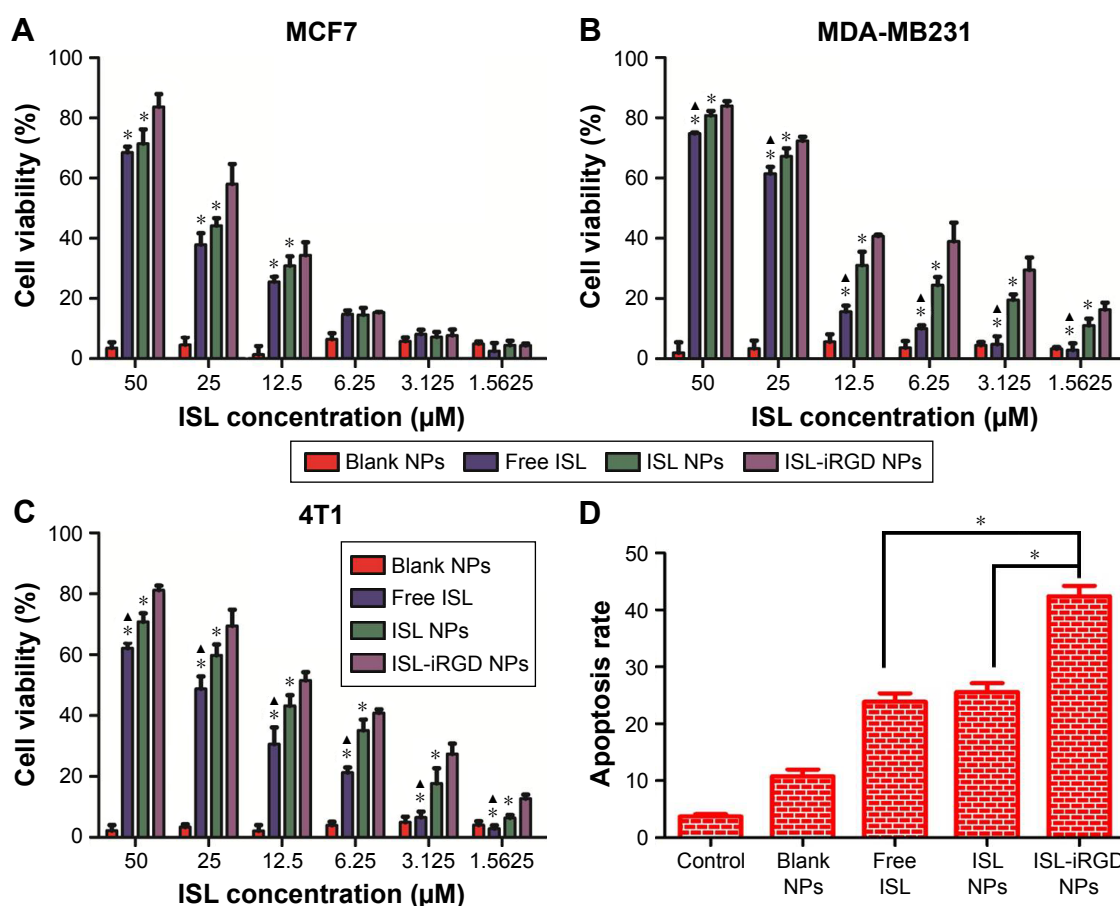
137.2 nm, which is also a good size range for passive targeting of NPs. Therefore, two methods were used to evaluate the cell-uptake ability of different formulas: fluorescence microscopy and FCM.

Fluorescence microscopy is a typical method for qualitative analysis of cell-uptake ability of NPs. The intensity of the dye in the image can reflect the concentration of target-drug uptake by certain cells indirectly. Figure 4A presents fluorescence-microscopy images of the different formulations taken up by MDA-MB231. The concentration of C_6 in all formulas was 85 ng/mL. After incubating with the different C_6 formulations for 4 hours, C_6 -iRGD NPs presented the greatest intensity and were most closely situated around

nuclei compared to C_6 NPs and free C_6 , which illustrated C_6 -iRGD NPs were prone to internalize in MDA-MB231 cells and showed excellent targeting ability.

Qualitative analysis of the NPs was measured by FCM to confirm uptake efficacy and evaluate the manner of uptake of C_6 -iRGD NPs. As shown in Figure 4B, after incubation of MDA-MB231 with different C_6 formulations (C_6 -iRGD NPs, C_6 NPs, and free C_6) for 4 hours, the FCM-detection profiles and histogram results of mean fluorescence intensity showed the intensity of C_6 in C_6 -iRGD NPs was strongest ($P < 0.05$), followed by C_6 NPs and free ISL. This confirmed the results of fluorescence microscopy, and also illustrated the strong cell-uptake ability of ISL-iRGD NPs. Moreover, it also indicated that the high uptake capacity of the system may have been due to its nanoscale and the iRGD peptide.

The manner of uptake of C_6 -iRGD NPs was also detected by FCM. The FCM profile and histogram both showed the mean intensity of C_6 increased with ISL dosage (Figure 4C) or time (Figure 4D and Figure S2), which indicated that the cell-uptake efficacy in MDA-MB231 of C_6 -iRGD NPs

**Figure 3** In vitro cytotoxicity and apoptosis analysis of ISL-iRGD NPs.

Notes: Cell viability of MCF7 (A), MDA-MB231 (B), and 4T1 (C) breast cancer cell lines after incubation with ISL-iRGD NPs, ISL NPs, blank NPs and free ISL for 24 hours. Apoptosis analysis of MDA-MB231 after treated by ISL-iRGD NPs, ISL NPs, blank NPs and free ISL (D). * $P < 0.05$ vs free ISL; $\Delta P < 0.05$ vs ISL NPs.

Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles.

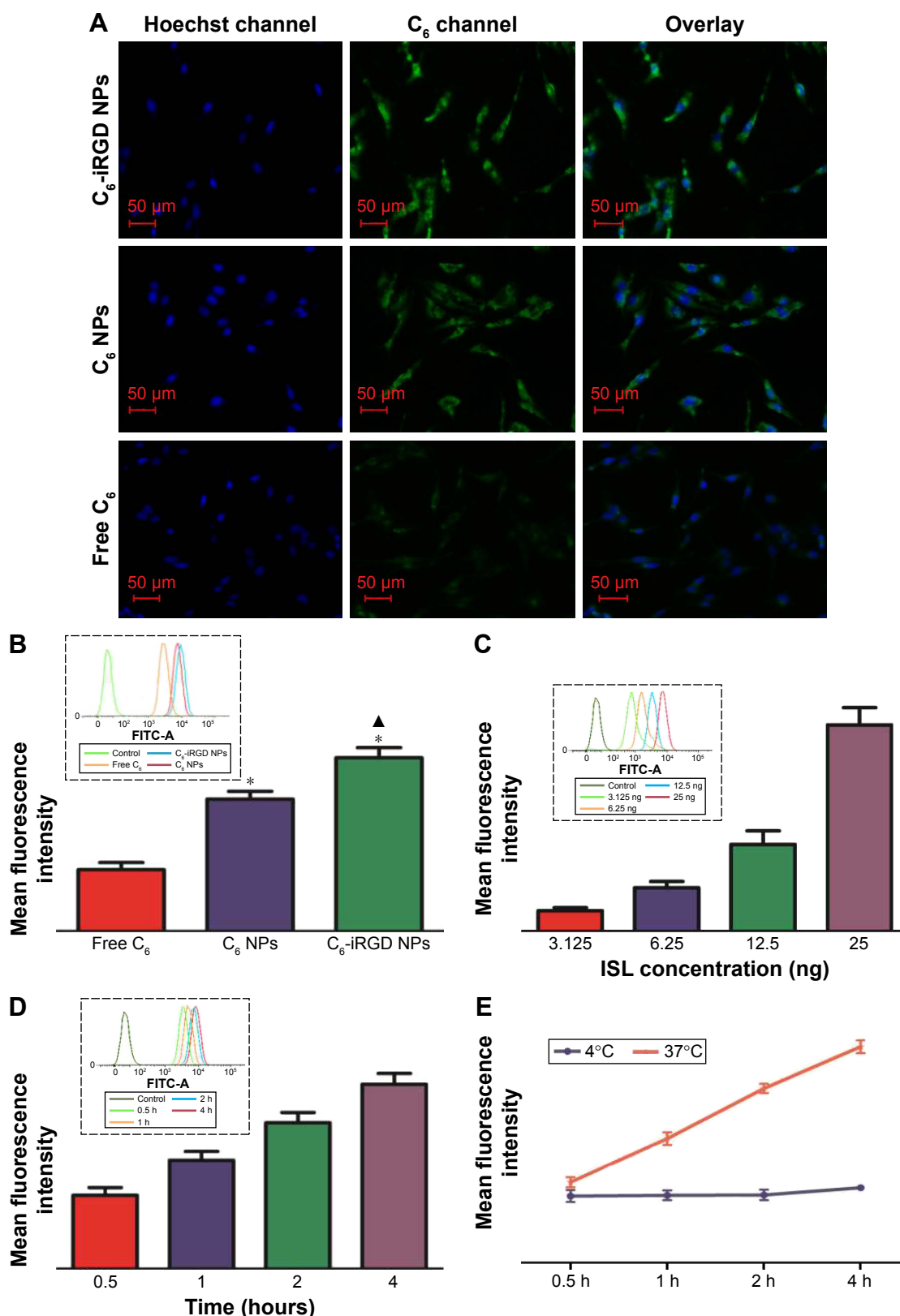


Figure 4 Cell-uptake evaluation of different C_6 formulations using the MDA-MB231 cell line.

Notes: (A) Fluorescent images of MDA-MB231 cells incubated with different C_6 formulations (C_6 -iRGD NPs, C_6 NPs, and free C_6) for 4 hours; magnification $\times 200$. C_6 incubation concentration was 85 ng/mL. (B) Quantitative measure of C_6 uptake in different C_6 formulations (C_6 -iRGD NPs, C_6 NPs, and free C_6) after incubation for 4 hours. C_6 -incubation concentration was 25 ng/mL. * $P < 0.05$ vs free C_6 ; $\Delta P < 0.05$ vs C_6 NPs. (C) Quantitative measure of C_6 uptake in C_6 -iRGD NPs at different dosages (25 ng/mL, 12.5 ng/mL, 6.25 ng/mL, and 3.125 ng/mL) after incubation for 4 hours. (D) Quantitative measure of C_6 uptake in C_6 -iRGD NPs at different incubation time points (0.5, 1, 2, and 4 hours). C_6 concentration was 25 ng/mL. (E) Mean fluorescence intensity versus time profile after incubation with C_6 -iRGD NPs at 4°C and 37°C for 4 hours.

Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles.

occurred in a dosage-dependent or time-dependent manner, respectively. Further analysis demonstrated that the intensity of C_6 at 37°C is stronger than at 4°C at all time points (Figure 4E), and intensity increased obviously with time at 37°C but not at 4°C, which suggested uptake behavior relied mainly on endocytosis, rather than physical adsorption or passive diffusion. In other words, cell uptake was energy-dependent.³⁰ This cell-uptake ability indicated that the iRGD peptide can enhance the cell-uptake efficacy of C_6 -iRGD NPs in time-dependent, dosage-dependent, and energy-dependent manners, which is the main reason for the excellent efficacy of ISL-iRGD NPs in anti-breast cancer, rather than ISL-NPs or free ISL.

In vivo antitumor efficacy of ISL-iRGD NPs

Based on the strong sensitivity of the 4T1 cell line on ISL-iRGD NPs, the classic 4T1-bearing nude-mouse model was selected to evaluate the in vivo anticancer efficacy of different formulations. Images of the tumors of the different groups are shown in Figure 5A. Further analysis comparing mean tumor volume between the ISL-iRGD NPs and the other three groups (free ISL, blank NPs, and control) showed significant difference ($P < 0.05$) was observed from the 14th day (Figure 5B). For the end point, the mean tumor volume of ISL-iRGD NPs decreased to about 474 mm³ compared to the other three groups (Figure S3). In addition, the mean tumor-volume burden also confirmed a similar outcome with mean tumor-volume analysis. The mean tumor burden in ISL-iRGD NPs was lower than for free ISL, blank NPs, and the control group ($P < 0.05$), and slightly lower than the ISL NPs (Figure 5C). These results and the graphs indicate that ISL-iRGD NPs showed better anti-breast cancer efficacy than the other four groups (ISL NPs, free ISL, blank NPs, and control).

Furthermore, H&E staining was used to record morphology changes among the different groups (Figure 5E). The quantity of mitotic-bodies in ISL-iRGD NPs and ISL NPs was higher than the other three groups, suggesting the viability of tumor cells was inhibited in ISL-iRGD NPs and ISL NPs in vivo. However, the blank NPs had similar morphology to the controls, which confirmed the low/absent anti-breast cancer effects of the nanomaterials.

For assessment of toxicity of the different formulas, body weight and H&E staining of organs were used. Body weight varying with time was recorded for 20 days, and there was no significant difference among the five groups (Figure 5D), which showed these five formulae had no obvious side effects on the animals. On the other hand, H&E staining of

lungs, livers, and kidneys is shown in Figure S4. For lungs, all of the groups had lung metastasis detected, which met the basic characteristics of the 4T1-bearing mice model. For livers and kidneys, there were no remarkable morphological changes observed among the groups. All in all, this evidence confirmed the fact that our NPs had no obvious toxicity to the mice in this research.

As for dosage, 50 mg/kg/day was an effective dose for free ISL in nude mice in our preliminary studies.^{24,31–33} However, in this project the effective dose was decreased to 25 mg/kg every 2 days in the 4T1-bearing nude-mouse model. Also, the administration method was intraperitoneal rather than IV injection, which was because the volume of the NPs was too large to use IV injections. In sum, ISL-iRGD NPs had excellent ability for anti-breast cancer targeting in vivo with relative nontoxicity.

Biodistribution characteristics of ISL-iRGD NPs

In vitro targeting ability of ISL-iRGD NPs was evaluated by fluorescence microscopy and FCM in the foregoing sections, with satisfactory results, meeting our expectations. However, these findings focused only on manner of uptake at the cellular level. For passive uptake of NPs caused by the EPR effect, only in vivo research can be used. Moreover, in vivo biodistribution evaluation can show the targeting ability of ISL-iRGD NPs objectively and directly. Therefore, the in vivo biodistribution characteristics of ISL-iRGD analysis were assessed using the IVIS Spectrum in vivo imaging system.

Figure 6 shows the results of the biodistribution analysis of ISL-iRGD NPs. After IV injection of free Cy5, it was observed that the Cy5 accumulated mainly in the liver and intensity decreased at a higher speed than in the other two groups during the 24 hours. Moreover, little fluorescence was detected in tumors at 12 and 24 hours. However, in Cy5-iRGD NPs, fluorescence was detected at all four time points with relatively high intensity. Compared with Cy5 NPs, the detectable fluorescence area in tumors of Cy5-iRGD NPs was larger. The fluorescence signals of different organs or tissues in each group showed that the intensity of Cy5 in tumors followed an ascending order: Free Cy5, Cy5 NPs, and Cy5-iRGD NPs ($P < 0.05$). In addition, Cy5 was mainly accumulated in the liver with stronger intensity compared to the other two groups ($P < 0.05$). All in all, based on biodistribution analysis in vivo, Cy5-iRGD NPs showed better tumor-targeting ability than free Cy5 and Cy5 NPs, perhaps due to the tumor-homing iRGD peptide and EPR effect.

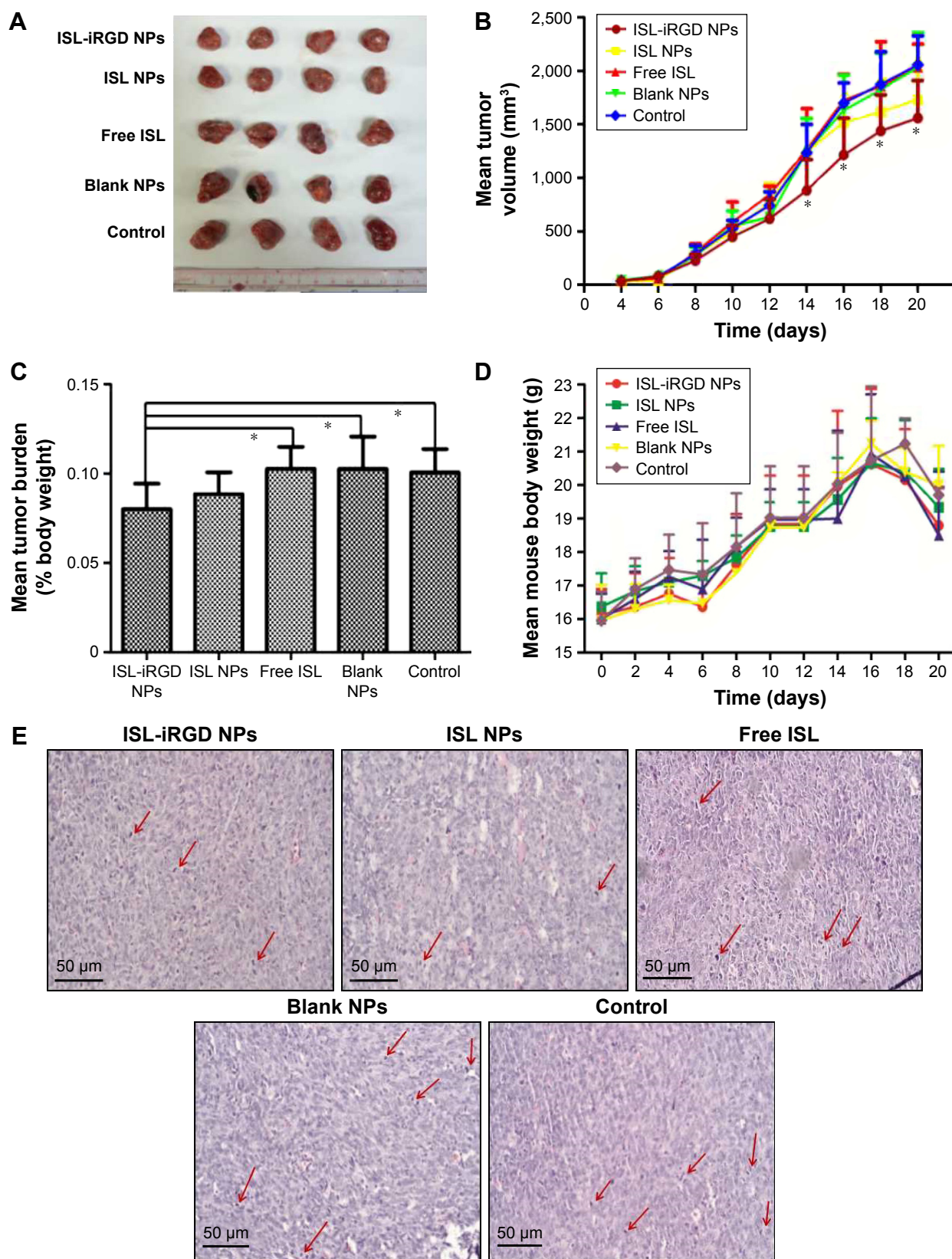


Figure 5 In vivo antitumor efficacy of ISL-iRGD NPs, ISL NPs, free ISL, blank NPs, and saline in the tumor-bearing mouse model.

Notes: (A) Graph of the tumor after intraperitoneal injection of different ISL formulations (25 mg/kg); (B) profile of mean tumor volume varying with time among different groups (* $P < 0.05$ vs free ISL, blank NPs, and saline); (C) mean tumor burden among different ISL formulations (* $P < 0.05$); (D) profile of the mean mouse body weight varying with time among different groups; (E) H&E staining on tumor (×20 objective). The red arrows indicate the mitotic-bodies of the cancer cells.

Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles; H&E, hematoxylin and eosin.

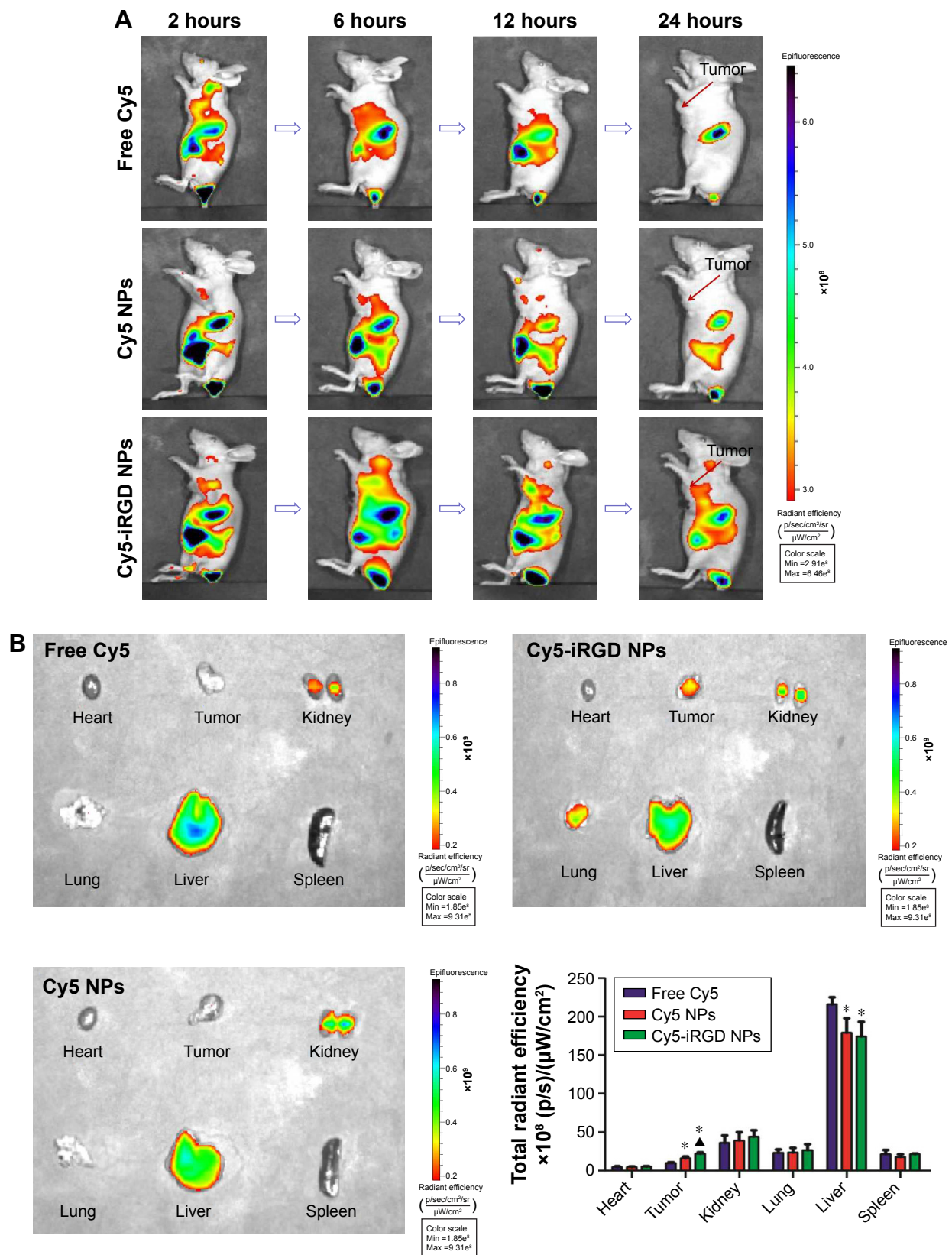


Figure 6 Biodistribution analysis of free Cy5, Cy5 NPs, and Cy5-iRGD NPs in the tumor-bearing mouse model.

Notes: Mice were injected intravenously via the tail vein with free Cy5, Cy5 NPs, and Cy5-iRGD NPs (35 $\mu\text{g}/\text{kg}$ mice). **(A)** Fluorescence images of mice treated with different Cy5 formulations in vivo at different time points (2, 6, 12, and 24 hours). **(B)** Fluorescence images of mice organs treated with different Cy5 formulations for 24 hours in vivo and fluorescent-signal analysis of different organs/tissues in each group. * $P < 0.05$ compared with free Cy5; $\Delta P < 0.05$ compared with Cy5 NPs.

Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles.

Conclusion

In this study, ISL-iRGD NPs were prepared successfully. The size of the NPs was about 130 nm, which is the appropriate size for the EPR effect in cancer therapy.²⁸ The ζ -potential of ISL-iRGD NPs was about -34 mV, which indicated a relatively stable system.²⁹ ISL-iRGD NP structure was also confirmed by four different methods, which demonstrated that the ISL was successfully encapsulated in the core of the NPs with high stability. In addition, they also had sufficient entrapment efficiency and loading efficiency. Compared with ISL NPs and free NPs, ISL-iRGD NPs had better efficacy in anti-breast cancer targeting and stronger targeting ability. In the pharmacodynamic study in vitro, ISL-iRGD NPs showed stronger inhibition effects and induced apoptosis effects. In the in vivo research, ISL-iRGD NPs showed stronger effects in inhibiting tumor growth and viability of tumor cells. The toxicity of the nanomaterials was also evaluated. The in vivo study outcome showed the nanomaterials had no significant damaging effects on normal tissue. The targeting ability of ISL-iRGD NPs was detected in vivo and in vitro. The data showed that ISL-iRGD NPs had better ability in increasing cell-uptake efficiency in a time-independent, dose-independent, and energy-independent manner. The biodistribution test also indicated that ISL-iRGD NPs were easier to target and accumulate in breast tumors. All in all, this new delivery system for encapsulating ISL was effective and can promote the development of ISL in breast cancer therapy in clinical practice.

The global burden of cancer has been increasing in the past few decades, raising the prospect of significant investment in health systems and thus posing a real medical problem. According to GloboCan data in 2012, breast cancer is the commonest cancer in women worldwide. Due to the serious toxicity of first-line synthetic agents, the high incidence of breast cancer and its correlation with cancer-related deaths worldwide have prompted more attention on discovering novel and effective alternatives with fewer side effects. Natural products, such as dietary phytochemicals, tea polyphenols, and resveratrol, are well recognized as effective agents in several human ailments, including cancer. A natural flavonoid, ISL has previously been demonstrated to exhibit antioxidant, anti-inflammatory, and tumor-suppression effects by inducing apoptosis, autophagy, and cell-growth inhibition. Nevertheless, the high lipophilicity and insufficient anticancer efficacy in comparison with synthetic agents like doxorubicin greatly limit its clinical efficacy. Recently, nanoscale

carriers have played a significant role in drug delivery. Various types of nanovehicles, such as liposomes, micelles, and nanocapsules, can encapsulate poorly soluble drugs to improve solubility. More importantly, anticancer drugs loaded in NPs can easily permeate into the leaky vasculature of tumor-tissue and control the drug-release behavior at the tumor site, which has been well described to enhance anticancer efficiency.

Lipid-related NPs, with liposomes as the classic type, and polymer-manufactured NPs, such as polymeric micelles and dendrimers, are the two dominant classes of nanoscale drug vehicles, with the increasing backing of approved drug products, clinical trials, and research experiments. Although many advantages of these nanocarriers have been demonstrated on occasion to improve solubility, body circulation, and selective tumor distribution, unfortunately both classes suffer limitations in terms of their physicochemical and biological properties. Specifically, most liposomes exhibit superior biocompatibility, biodegradability, and nonimmunogenic response, because natural lipids are analogs of biological membranes. However, liposomes display poor storage stability, and are easily cleared by the reticuloendothelial system. On the other hand, polymeric NPs have advantages in particle size, stability, and drug loading. Therefore, herein we employed lipid-polymer hybrid NPs constituted of PLGA, lecithin, and PEG to combine the advantages of lipid NPs and polymeric NPs. Moreover, iRGD peptides were grafted to DSPE-PEG_{2,000}-Mal to obtain tumor-targeting by the conjugation of the thiol group of iRGD to the Mal group of PEG.³⁰ Our research confirmed significant targeting ability and anti-breast cancer effects both in vivo and in vitro. How to synthesize such materials using simpler and cost-effective methods would be the key question, which requires addressing.

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Disclosure

The authors report no conflicts of interest in this work.

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Supplementary materials

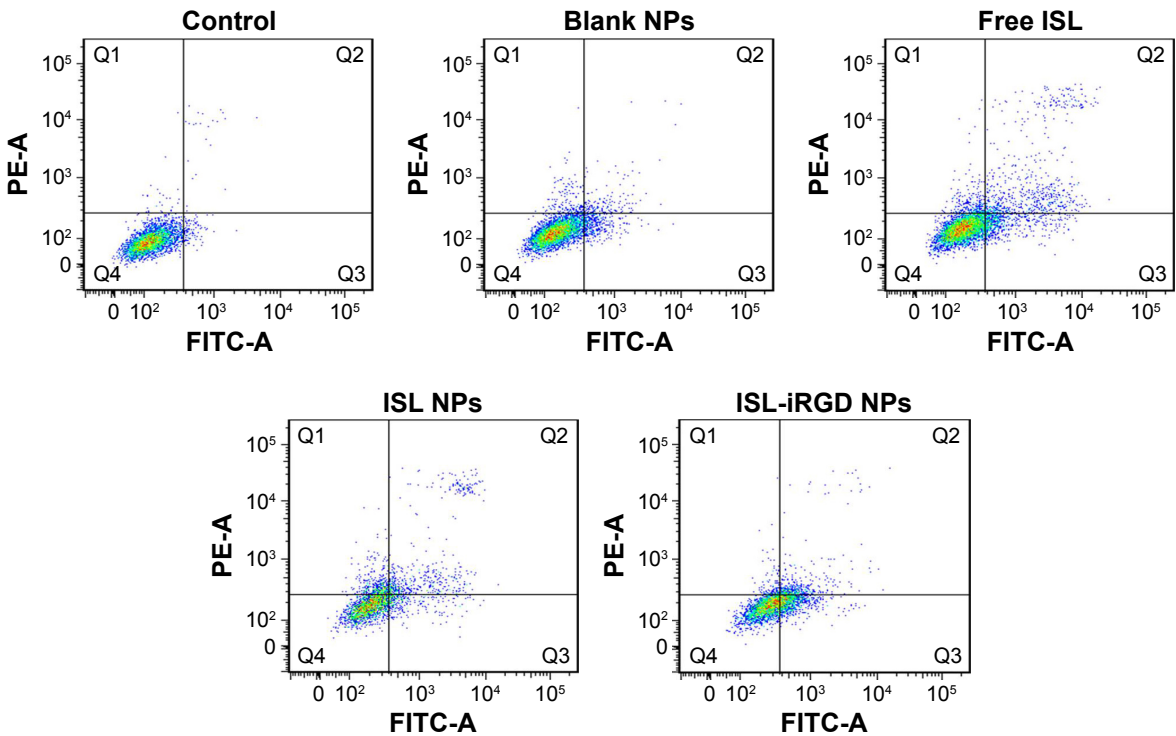


Figure S1 Flow cytometry of different ISL formulations for apoptosis analysis in MDA-MB231 cell lines.
Notes: Top left (Q1), necrotic cells (annexin V-FITC-PI⁺); top right (Q2), late apoptotic cells (annexin V-FITC-PI⁺); bottom left (Q3), live cells (annexin V-FITC-PI⁻); bottom right (Q4), early apoptotic cells (annexin V-FITC-PI⁻).
Abbreviations: ISL, isoliquiritigenin; PI, propidium iodide; NPs, nanoparticles.

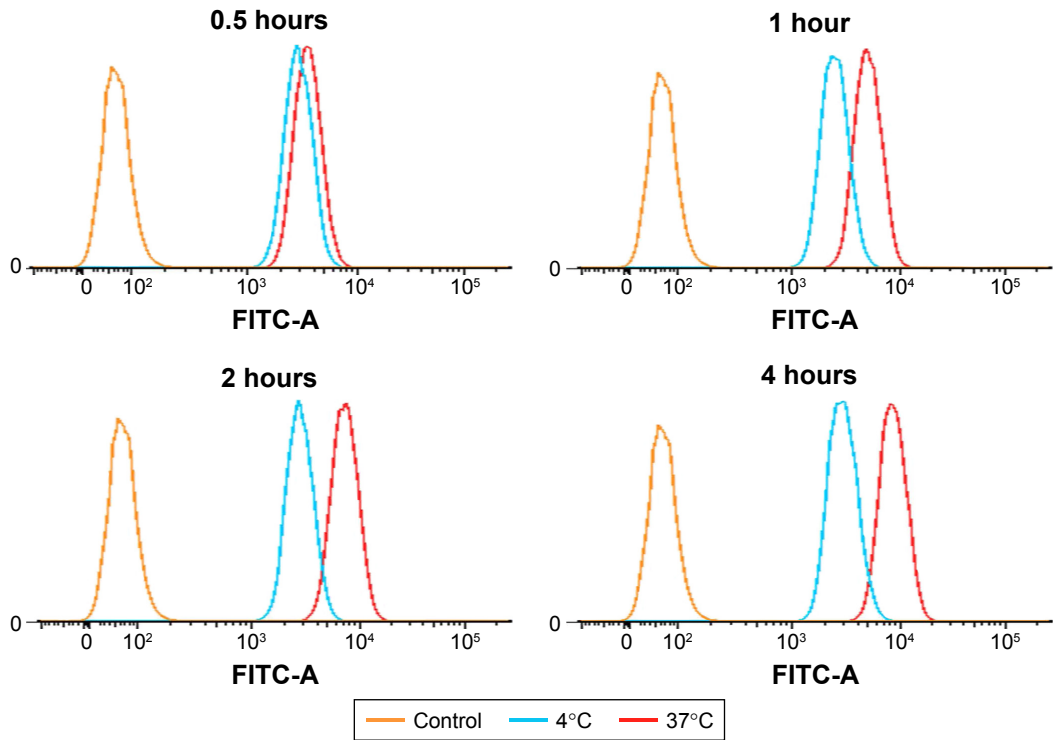


Figure S2 Flow-cytometry profile of MDA-MB231 cell line incubated with C₆-iRGD NPs at different incubation time points (0.5, 1, 2, and 4 hours) at 4°C and 37°C.
Abbreviation: NPs, nanoparticles.

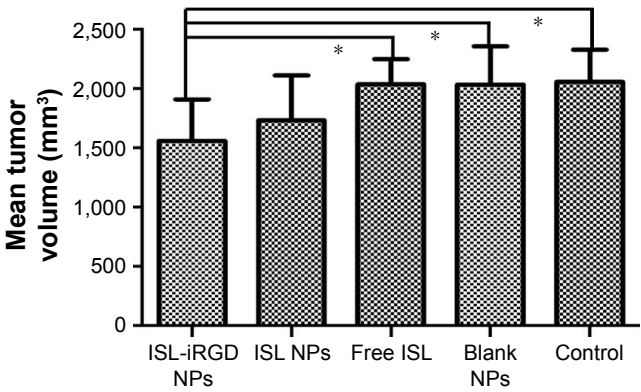


Figure S3 Mean tumor volume differences among different groups (* $P<0.05$).
Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles.

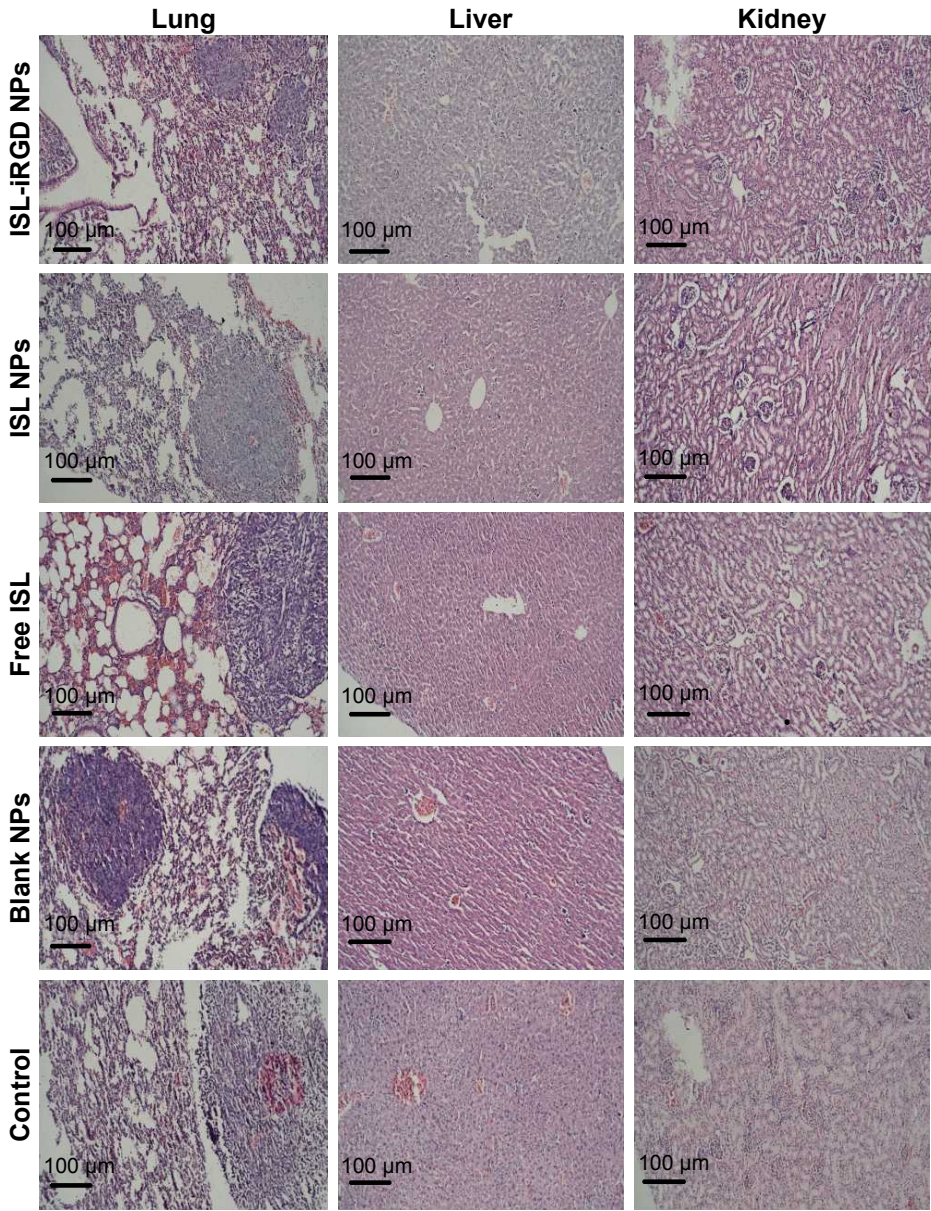


Figure S4 Hematoxylin and eosin staining on lung, liver, and kidney (10× objective).
Abbreviations: ISL, isoliquiritigenin; NPs, nanoparticles.

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